SYNTHESIS OF RACEMIC 11,12-CYCLOPROPYL ANALOGS OF HEPOXILINS A3 AND B3

Peter M. Demin¹ and Cecil R. Pace-Asciak^{*1,2}

¹Research Institute, Hospital for Sick Children, Toronto, Canada M5G 1X8 and ²Department of Pharmacology, University of Toronto, Toronto, Canada M5S 1A8

Summary: Stable biologically active racemic 11,12-cyclopropyl analogs of hepoxilins A₃ and B₃ were prepared via polyacetylenic intermediates.

Hepoxilins (Hx) A₃ and B₃ are naturally occurring biologically active metabolites of 12(S)-HPETE.¹ They have been shown to stimulate insulin secretion,^{2a,2b} Ca²⁺ transport across membranes^{2c} and to mobilize Ca²⁺ from intracellular stores in human neutrophils.^{2d} Additional studies have shown that HxA₃ increases vascular tone^{2e,2f} and vascular permeability.^{2g} HxA₃ appears to affect second messenger systems in the neutrophil releasing arachidonic acid and diacylglycerol.^{2h} Hepoxilins have also been implicated in neurotransmission with actions on mammalian²ⁱ and Aplysia neurons.^{2j} HxA₃ is relatively unstable biologically and chemically and can be metabolized into the corresponding trioxilins³ or peptido-hepoxilins⁴ by enzymatic mechanisms, the latter also possessing biological activity.^{2k}

Total chemical synthesis of native hepoxilins has been reported by several laboratories,^{5,6} which has afforded sufficient material for biological testing. In this connection, structural analogs of Hx which selectively modulate Hx actions may provide compounds through which the putative role of Hx may be investigated. Our approach has been to modify the unstable allylic epoxide group of HxA_3 through replacement with a cyclopropyl group to render it chemically and metabolically stable.

We describe herein the synthesis of stable racemic 11,12-cyclopropyl analogs of hepoxilins: methyl 8hydroxy-11,12-cyclopropyl-5(Z),9(E),14(Z)-eicosatrienoate (Δ HxA₃) and methyl 10-hydroxy-11,12cyclopropyl-5(Z),8(Z),14(Z)-eicosatrienoate (Δ HxB₃) based on the acetylenic approach to eicosanoids.^{6b,7}

1-Hydroxyundeca-2(E)-en-5-yne 1, which serves as key intermediate in the synthesis of native hepoxilin B series^{6a,b} is employed herein to synthesize Hx analogs. It was prepared as described.^{6b} 1 was treated with CH₂I₂ and Zn-Cu couple in dry ether giving racemic ($2S^*, 3S^*$)-2,3-cyclopropylalcohol 2; the latter was then oxidized to aldehyde 3 with pyridinium dichromate (see Scheme). Two subsequent condensations of aldehyde 3 with Li-derivative of propargyl chloride led to cyclopropylcarbinol 4. ¹H-NMR spectrum showed a 7:3 ratio between two diastereomers. Chloride 4 was reacted with methyl hexynoate in the presence of equimolar amounts of CuI and NaI in DMF resulting in the triacetylenic analog of Δ HxB₃ 5 obtained as a unseparable mixture with the same epimeric ratio. Selective Lindlar hydrogenation of triacetylene 5 and separation by HPLC gave two C10-epimers of more and less polar Δ HxB₃ methyl esters



^aCH₂I₂, Zn(Cu), ether, 1h, reflux. ^bPy₂H₂Cr₂O₇, CH₂Cl₂, 30 min, 20^oC. ^cHC=CCH₂Cl, n-BuLi, ether, -78^oC, 15 min, than H₂O. ^dHC=C(CH₂)₃COOMe, CuI,NaI, K₂CO₃, DMF, 10 h, 20^oC. ^eH₂, Pd/Pb/CaCO₃, C₆H₆, quinoline. ^fAc₂O, Py, 10 h, 20^oC. ^gPdCl₂(MeCN)₂, THF, 3 h, 20^oC. ^hNaOH, MeOH/H₂O (2:1), 5 h, 20^oC. ⁱCH₂N₂, ether

6a,b in 7:3 ratio as a colorless oil. The same ratio between more and less polar epimers was obtained for native HxB₃ methyl esters when similar condensations of appropriate aldehyde with Li-derivative of terminal acetylene were used.^{6b,7} On this basis we expected the same relative configuration for more and less polar Δ HxB₃ as for native HxB₃. This proposition was confirmed by the consideration of NMR spectra of individual Δ HxB₃ methyl esters 6a. 6b. NMR spectra had shown larger coupling constant J_{10,11} for more polar epimer (d, 7.8 Hz) than for less polar epimer (d, J 7.3 Hz).⁸ These data were in agreement with those described for α , β -cyclopropylcarbinolic systems.⁹ Also oxidation of both fully saturated Δ HxB₀ (Δ HxB₀) epimer and native saturated HxB (HxB₀) by pyridinium dichromate into a corresponding ketone followed by treatment of the latter with sodium borohydride resulted in a mixture of initial α , β -cyclopropyl- and α , β -epoxycarbinols, respectively in 1.7:1 ratio between less and more polar epimers in both cases. This ratio was similar to that described for reduction of α , β -epoxyketones to α , β -epoxycarbinols by NaBH4 with preference of *erythro*epoxycarbinol.¹⁰ On this basis, we concluded that the more polar Δ HxB₃ epimer 6a has *threo* or *syn* or (10S*,11R*,12S*)-configuration and the less polar epimer of Δ HxB₃ 6b has *erythro* or *anti* or (10R*,11R*,12S*)-configuration.¹¹

To obtain the two C8-epimeric Δ HxA₃ methyl esters 7a,b we used the stereo controlled rearrangement of allylic acetates catalyzed by Pd(II).¹² Treatment of individual Δ HxB₃ acetates 8a, 8b obtained from 6a, 6b with 0.1 eqv. of PdCl₂(MeCN)₂ in THF led to the mixtures of Δ HxB₃ and Δ HxA₃ acetates 8a, 9a and 8b, 9b in ca. 1:1 ratio in both cases which could be separated from each other by HPLC.¹³ Following hydrolysis, two individual C8-epimers of Δ HxA₃ 7a, 7b were obtained. On the basis of an S_N2' reaction mechanism¹² we refer to the more polar Δ HxA₃ (obtained from *anti* Δ HxB₃) as *syn* or (8R*,11S*,12S*)-epimer 7b, and less polar Δ HxA₃ as *anti* or (8S*,11S*,12S*)-epimer 7a, respectively. The chromatographic properties of Δ HxA₃ methyl esters 7a, 7b were also similar to native HxA₃ methyl esters with known relative configuration.¹⁴

Preliminary biological testing has shown that the more polar isomers of Δ HxA₃ and Δ HxB₃ dosedependently (0.05-0.5µg/ml) inhibit the rise in free intracellular Ca²⁺ in human neutrophils evoked by 3µg/ml of HxA₃.¹⁵ Details of these studies will be reported fully at a later time.

References and Notes

1. C.R. Pace-Asciak, J. Biol. Chem. 259: 8332-8337 (1984).

Biological activities of HxA3 and HxA3-C: (a) C.R. Pace-Asciak, and J.M. Martin, Prostagl. Leuk. Med.
16: 173-180 (1984); (b) C.R. Pace-Asciak, J.M. Martin, E.J. Corey, W-G. Su, Biochem. Biophys. Res. Comm.
128: 942-946 (1985); (c) L.O. Derewlany, C.R. Pace-Asciak and I.C. Radde, Can. J. Physiol. Pharmacol. 62:
1466-1469 (1984); (d) S. Dho, S. Grinstein, E.J. Corey, W-G. Su, and C.R. Pace-Asciak, Biochem. J. 266: 63-68 (1990); (e) O. Laneuville, R. Couture, and C.R. Pace-Asciak, Br. J. Pharmacol. 105: 297-304 (1992); (f) O. Laneuville, R. Couture, and C.R. Pace-Asciak, Br. J. Pharmacol. 107: 808-812 (1992); (g) O. Laneuville, E.J. Corey, R. Couture, and C.R. Pace-Asciak, Eicosanoids 4: 95-97 (1991); (h) S. Nigam, S. Nodes, G. Cichon, E.J. Corey, and C.R. Pace-Asciak, Biochem. Biophys. Res. Commun. 171: 944-948, 1990; (i) P.L. Carlen, N. Gurevich, P.H. Wu, W-G. Su, E.J. Corey, and C.R. Pace-Asciak, Brain Res. 497: 171-176, 1989; (j) D. Piomelli, E. Shapiro, R. Zipkin, J.H. Schwartz, and S.J. Feinmark, Proc. Natl. Acad. Sci. USA 86: 1721-1725 (1989); (k) C.R. Pace-Asciak, O. Laneuville, W-G. Su, E.J. Corey, N. Gurevich, P. Wu, and P.L. Carlen, Proc. Natl. Acad. Sci. USA 87: 3037-3041 (1990).

3. C.R. Pace-Asciak, and W-S. Lee, J. Biol. Chem. 264: 9310-9313 (1989).

4. C.R. Pace-Asciak, O. Laneuville, M. Chang, C.C. Reddy, W-G. Su, and E.J. Corey, Biochem. Biophys. Res. Commun. 163: 1230-1234 (1989).

Syntheses of HxA₃: (a) E.J. Corey and W.-G. Su, *Tetrahedron Lett.* 25, 5119-5122 (1984); (b) P. Chabert,
Mioskowski and J.R. Falck, *Tetrahedron Lett.* 30, 2545-2548 (1989); (c) P.M. Demin, L.L. Vasil'eva,
Yu.Yu. Belosludtsev, G.I. Myagkova, and K.K. Pivnitskii, *Bioorg. Khim.* 16: 571-572 (1990), *Chem. Abstr.* 113: 131804q; (d) S. Lumin, J.R. Falck, J. Capdevila and A. Karara, *Tetrahedron Lett.* 33: 2091-2094 (1992).

6. Syntheses of HxB₃: (a) E.J. Corey, J. Kang, B.C. Laguzza and R.L. Jones, *Tetrahedron Lett.* 24, 4913-4916 (1983); (b) P.M. Demin, L.L. Vasil'eva, M.A. Lapitskaya, Yu.Yu. Belosludtsev, G.I.Myagkova, and K.K. Pivnitskii, *Bioorg. Khim.* 16: 127-128 (1990), *Chem. Abstr.* 113: 40257x; (c) W.-L. Wu and Y.-L. Wu, J. *Chem. Soc. Perkin Trans.* (1) 2705-2707 (1992).

7. Yu.Yu. Belosludtsev, G.I. Myagkova, R.P. Evstigneeva, N.I. Bobrova, and K.K. Pivnitskii, *Bioorg. Khim.* 13: 1125-1131 (1986), *Chem. Abstr.* 109: 54511e.

8. Chromatographic and spectral data for 6a: $R_f 0.39$ (C₆H₆-Et₂O, 85:15, 3 developments). Mass-spectrum, t-BDMS-derivative (m/z, % of related intensity): 462 ([M]⁺, 0.06), 431 ([M - OMe]⁺, 0.80), 405 ([M - t-Bu]⁺, 37), 324 ([C1-C11]⁺, 20), 211 (12), 169 (12), 105 (21), 75 (100). 6b: $R_f 0.51$ (C₆H₆-Et₂O, 85:15, 3 developments). Mass-spectrum, t-BDMS-derivative (m/z, % of related intensity): 462 (0.04), 431 (0.35), 405 (20), 334 (4.7), 324 (2.8), 215 (4.2), 211 (3.0), 169 (6.0), 105 (26), 75 (100). 1H-NMR spectra (500 MHz) show the difference in signals belong to protons at chiral groups as follows (δ , ppm): 6a: 0.39 (dt, 1H, J 4.8 and 8.1 Hz, cyclopropyl-H), 0.53 (dt, 1H, J 4.8 and 8.5 Hz, cyclopropyl-H), 0.68 (m, 1H, H¹¹), 0.83 (m, 1H, H¹²), 3.95 (ddd, 1H, J 1.0, 7.8 and 7.8 Hz, H¹⁰). 6b: 0.33 (dt, 1H, J 5.1 and 8.4 Hz, cyclopropyl-H), 0.81 (m, 2H, H¹¹ + H¹²), 3.97 (ddd, 1H, J 3.0, 7.3, and 7.3 Hz, H¹⁰).

9. G. Descotes, A. Menet, and F. Collonges, Tetrahedron 29: 2931-2935 (1973).

10. J.L. Pierre, and P. Chautemps, Tetrahedron Lett. 4371-4374 (1972).

11. The absolute configuration of the carbinolic centre of cyclopropyl analogs of HxB_3 is opposite to configuration of native HxB_3 while the relative configuration remains the same.

12. P.A. Grieco, T. Takigava, S.L. Bongers, and H. Tanaka. J. Am. Chem. Soc. 102: 7587-7588 (1980).

13. ¹H-NMR spectra for acetates **9a,9b** were identical to each other (500 MHz, δ , ppm): 0.57 (m, 2H, cyclopropyl-H), 0.80, 1.15 (m, 2H, H¹¹ and H¹²), 0.88 (t, 3H, J 6.91 Hz, H²⁰), 1.28-1.32 (m, 6H, H¹⁷ + H¹⁸ + H¹⁹), 1.69 (quintet, 2H, J 7.4 Hz, H³), 1.99-2.07 (m, 6H, H² + H⁴ + H¹⁶), 2.01 (s, 3H, OAc), 2.32-2.37 (m, 4H, H⁷ + H¹³+H¹³'), 3.67 (s, 3H, COOMe), 5.19 (dt, 1H, J 6.7 and 6.8 Hz, H⁶), 5.29 (dd, 1H, J 8.7 and 15.5 Hz, H¹⁰), 5.38 (m, 3H, H⁵ + H⁹ + H¹⁴ + H¹⁵), 5.44 (m, 1H, H⁸).

14. Chromatographic and spectral data for **7a**: $R_f 0.46$ (C_6H_6 -Et₂O, 85:15, 3 developments), mass-spectrum, t-BDMS-derivative (m/z, % of related intensity): 431 ([M - OMe]+, 0.18), 405 ([M - t-Bu]+, 4.7), 351 ([C¹-C¹²]+, 0.18), 321 ([C⁸-C²⁰]+, 100), 197 (62), 189([C⁸-C²⁰] - t-BuMe₂SiOH, 17], 171 (27), 75 (79), 73 (83). **7b**: $R_f 0.50$ (C_6H_6 -Et₂O, 85:15, 3 developments), mass-spectrum, t-BDMS-derivative (m/z, % of related intensity): 431 (0.06), 405 (1.0), 351 (0.04), 321 (28), 197 (24), 189 (7.3), 171 (12), 75 (56), 73 (100).

15. This research has been funded by grants to CRP-A from the MRC of Canada and from ZymoGenetics, USA.

(Received in USA 13 April 1993; accepted 11 May 1993)